

Genomic Analysis of fgf in zebra Fish (Danio Rerio)

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Mini Review

Abstract

FGFs are abbreviated as "Fibroblast growth factors". These "FGFs" are the "secreted" development factors of polypeptide. The "signaling" scheme of FGFs is actually the vital part which "plays" a significant role in many processes of development in vertebrates. The humans also have FGFs family. This family comprises of 22 members. On the other hand, zebra fish family has 16 members while eleven more zebra fish fgf were identified by homo-logy based search. This search was conducted in zebra fish genome and cDNA databases. By calculating additional member's fgf family of zebra fish now consist of 27 members at least. By the conduction of polygenetic processes and gene location analyses, we identified relationships between human FGFs genes and zebra fish Fgf genes. This paper also evaluates and represents the phylogenetic information of fgf zebra fish family.

Keywords: Fgf, zebra fish, Phylogenetic analysis, Human fgf, Gene location, Fibroblast

Introduction

(FGFs) are actually the development factors named as "Fibroblast development factors". They structure a group of commonly "extracellular" flagging "peptides", which are the controllers of various "organic" cycles going from "cell-multiplication" to the command of undeveloped advancement in "metazoans" [1]. Most FGF qualities are discovered dissipated all through the "genome". In human, "22 FGF" qualities have been recognized and the chromosomal areas of all with the exception of FGF16 are known. A few human FGF qualities are bunched inside the "genome" [2]. "FGF3, FGF4 and FGF19" are situated on "chromosome 11q13" and are isolated by just forty and ten kb, separately; "FGF6" and " FGF23" are situated inside fifty kb on " chromosome" [3] "12p13; and FGF17 and FGF20" guide to "chromosome 8p21-p22". These quality areas demonstrate that the "FGF-quality-family" was produced by "quality" and "chromosomal" duplication and movement throughout advancement. [4] Curiously, a transcriptionally dynamic segment of human FGF7, situated on chromosome "15q13-q22", has been intensified to around 16 duplicates, which are scattered all through the human genome [4]. "FGFs" are indicated in the two "vertebrates" and "spineless" creatures [5]. Various "FGF" qualities are distinguished in "vertebrates", incorporating 10

"FGFs" in zebra fish "FGF2-4, 6, 8, 10, 17a, 17b, 18, 24", 6 in "Xenopus" "FGF2-4, 8-16", thirteen in "chickens" "FGF1-4, 8-10, 12, 13, 16, 18-22", 22 in "mice" "FGF1-18, 20-23" and people "FGF1-14, 16-23", though just three "Drosophila" "FGF" qualities [6] and two "Caenorhabditis" elegans "FGF" qualities have been seen in "spineless" creatures [7]. Human "FGFs" contain twenty-two individuals: "FGF1, FGF2, FGF3 (INT2), FGF4, FGF5, FGF6, FGF7 (KGF), FGF8 (AIGF), FGF9, FGF10, FGF11, FGF12, FGF13, FGF14, FGF16, FGF17, FGF18, FGF19, FGF20, FGF21, FGF22, and FGF23" [8]. The FGF family contains twenty-three individuals, despite the fact that there is just eighteen "FGFR" ligand. Four relatives don't tie with "FGFR" as "FGF" "homologous" "elements" ("FGF11, FGF12, FGF13, and FGF14") [9] and are all the more accurately alluded to as "FGF-homologous-variables" [10]. Furthermore, there is no human "FGF15" quality; the quality "orthologous" to "mouse" "FGF15" is "FGF19" [11]. By "phylogenetic" examination, the "human FGF" quality family can be separated in to 7 sub families: "FGF1, FGF4, FGF7, FGF8, FGF9, FGF11, and FGF19. The FGF1, FGF4, FGF7, FGF8, FGF9, FGF11, and FGF19 sub families include FGF1 and 2, FGF4, 5, and 6, FGF3, 7, 10, and 22, FGF8, 17, and 18, FGF9, 16, and 20, FGF11, 12, 13, and 14, and FGF19, 21, and 23", separately [12]. As opposed to "phylogenetic" investigation, quality area examination shows that the human FGF quality family can be isolated in to 6 sub families: "FGF1/2/5, FGF3/4/6/19/21/23, FGF7/10/22, FGF8/17/18, FGF9/16/20, and FGF11/12/13/14" [13]. Individuals from the "FGF8, FGF9, and FGF11" sub-families are familiar to those of the "FGF7/10/22, FGF8/17/18, FGF9/16/20, and FGF11/12/13/14" sub families in the quality area investigation [14]. During undeveloped turn of events, FGFs have assorted jobs in directing cell expansion, relocation and separation [15]. In the grown-up life form, FGFs are homeostatic factors and capacity in tissue fix and reaction to injury. At the point when improperly communicated, some FGFs can add to the pathogenesis of malignant growth [16]. A subset of the FGF family, communicated in grown-up tissue, is significant for neuronal sign transduction in the focal and fringe sensory systems. FGFs are little proteins (somewhere in the range of 17 and 34 kDa)

described by a generally very much preserved focal space of "120 to 130 amino acids" [17]. This area is coordinated into twelve "antiparallel" "β" sheets shaping a three-sided substance called "beta-trefoil" [18] When all is said in done, FGFs work through restricting to a "tyrosine-kinase-receptor" "FGFR" on the outside of the cell film. Two "FGF-ligands" tie a "dimeric-receptor" within the sight of "heparin-sulfate-proteoglycan" permitting the "transphosphorylation" and attachment of the "intracellular-tyrosine-kinase" "space of the receptor" [2]. Through the initiation of the "cytoplasm" pathways, the "FGF-signal-controls" a few significant cell capacities like cell expansion, relocation, separation, or endurance [9].

Material and Method

Identification of fgf genes in zebra fish

PSI Blast, protein blast [BLAST P] and Delta Blast were performed in zebra fish to regain the protein and cDNA sequence of fgf genes. After it, by using the different databases, the target sequences were processed. For example: Pfam databases, smart databases, and NCBI [conserved] data bases. cDNA sequences, chromosomal location, length of protein and their genomic information [genomic information related to the fgf genes] were taken from NCBI.

Gene structure and prediction:

Gene structure display server is a freely online available Server. Schematic representation and gene structure analysis is done by this server.

Gene location analysis

Ensemble genome browser (ensembl.org) is a browser by which the positions of "human" and "zebra fish" "genome" on "chromosomes" are obtained. However, "psychogenetic" analyses and "gene located" analyses pointed the potential

evolutionary relationship. However, gene evolutionary relationships can never be determined only by it. However, the "psychogenetic" analyses and gene located analyses also indicate potential evolutionary relationship in gene formation [19]. At human "fgf7" and "fgf 22" loci and zebra fish "fgf7" and "fgf22" loci, conserved gene orders were examined. They indicate that zebra fish "fgf7" and "fgf22" are orthologues of human "fgf7" and "fgf22". At human "fgf10" and "fgf10a" loci a "conserved" gene-order was obtained. It points that "fgf10a" is a "zebra fish" of "human-fgf10"

Phylogenetic analysis

An online software tool named as mega 7 was used to make Fgf genes in zebra fish of polygenetic analysis. it is accessed from(<https://mega.software.informer.com/7.2/>) However "polygenetic analysis" points that human "Fgf gene family" are further termed in to sub families. Such as "FGF1, FGF4, FGF7, Fgf8, Fgf9 and Fgf11". These subfamilies consist of "Fgf1=2, fgf4=5, fgf3=7, fgf10=22, fgf8=17=18, fgf9=16=20, fgf11=12=13=14 and fgf19=21=23" by order. On the contrary, "zebra fish" "fgf-gene-family" are also further termed in to sub families. Zebra fish "fgf" sub families are potentially coherent with the human fgf families. Human "FGF5" is a fellow member of the "FGF4" sub family, and "zebra fish fgf5" is a member of the fgf1 subfamily.

Results

Phylogenetic analysis

Figure 1 to 3

Discussion

FGFs abbreviated as fibroblast growth factors. The humans also have FGFs family. This family comprises of 22 members [20]. On the other hand, zebra fish



Figure 1. The evolutionary history was inferred using the Neighbor-Joining method.

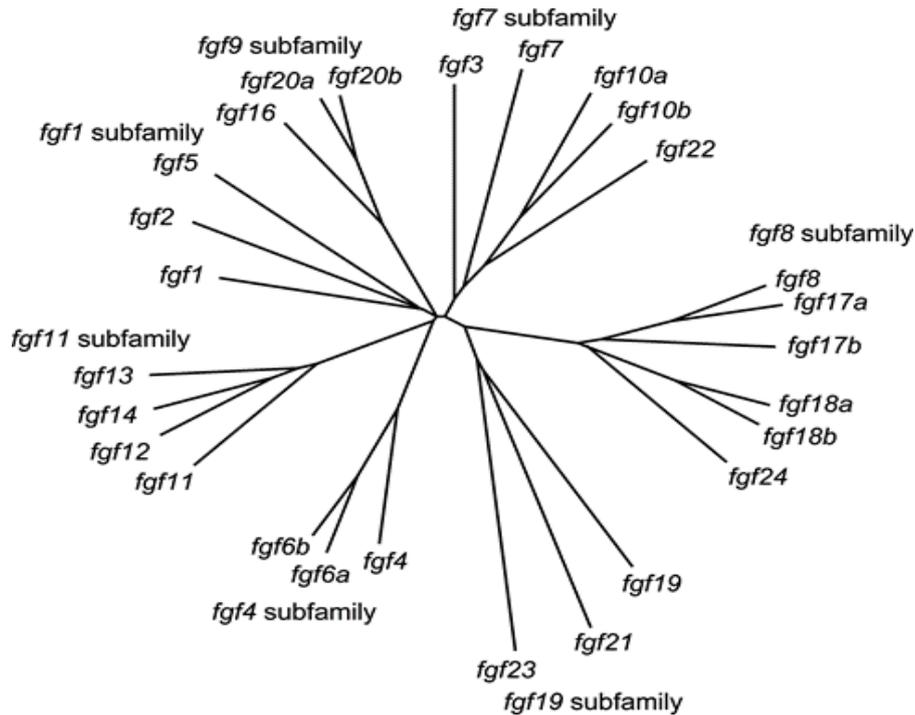


Figure 2. Evolutionary relationship of fgf gene family in zebra fish.

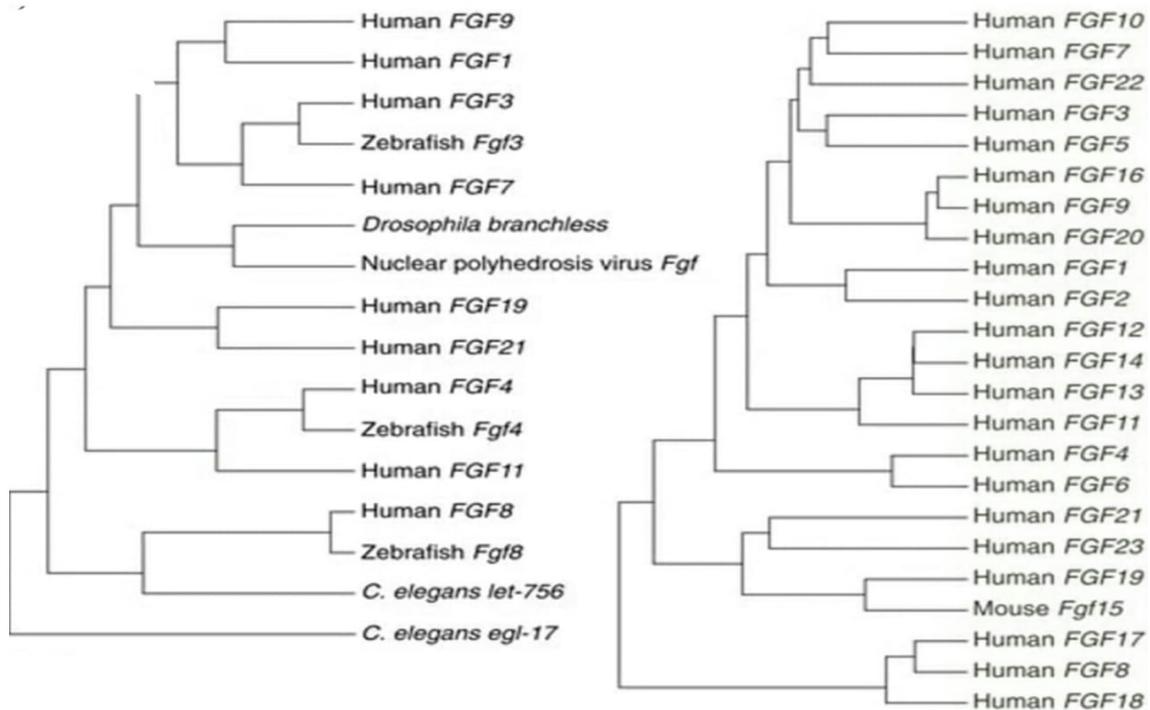


Figure 3. left side of the image shows evolutionary relationships between vertebrates, and invertebrates whereas the right side of image shows evolutionary relationships of murine and human fgf.

family has 16 members while eleven more zebra fish fgfs were identified by homo-logy based search. This search was conducted in zebra fish genome and cDNA databases. By calculating additional members FGF family of zebra fish now consist of 27 members at least [21]. By the conduction of "polygenetic "processes and "gene location analyses "we pointed relationships between human "fgf-genes" and zebra

fish "Fgf-genes" [22]. After the research, we found that there were no extra undefined fgf. During the evolution, it might be possible that fgf9 has been lost. In zebra fish fgf family, there are six paralog [23]. If there is a comparison of mammalian gene with Teleost fish, teleost was shown in it. Including zebra fish there are more often to homo- logos of mammalian equivalent. It proposed that that

shortly after the teleost radiation, there is additional genome duplication [24]. This may be partially or wholly genome duplication. It is preceded by rapid-gene-loss. It is because gene "duplication" works only for the 20% of zebra fish genes identity. By the process of gene duplications, zebra fish fgf paralog were generated. This whole process is done by the location analyses of zebra fish fgf genes [25]. Fibroblast development factors (FGFs) are a group of fundamental factors that control creature advancement and physiology; in any case, their job in cetacean advancement isn't obviously perceived. Here, we sequenced the blade whale genome and dissected FGFs from 8 cetaceans. FGF22, a hair follicle-advanced quality, displayed pseudogenization, demonstrating that the capacity of this quality is not, at this point fundamental in cetaceans that have lost the majority of their body hair. A transformative investigation uncovered marks of positive determination for FGF3 and FGF11, qualities identified with ear and tooth advancement and hypoxia, separately. We discovered a D203G replacement in cetacean FGF9, which was anticipated to influence FGF9 homodimerization, recommending that this quality assumes a part in the procurement of inflexible flippers for productive moving. Cetaceans use low bone thickness as a lightness control system, yet the fundamental qualities are not known. We tracked down that the declaration of FGF23, a quality related with diminished bone thickness, is extraordinarily expanded in the cetacean liver under hypoxic conditions, subsequently embroiling FGF23 in low bone thickness in cetaceans. Out and out, our outcomes give novel bits of knowledge into the parts of FGFs in cetacean variation to the oceanic climate.

Furthermore, the "phylogenetic analyses" and "gene located analyses" pointed that orthologues of "fgf6b" and "fgf6a" are the "paralog" of "fgf6b". "fgf10a" was basically examined as "orthologues" of human "fgf10". "Fgf10b" is a "paralog" of "fgf10" [26]. Zebra fish "fgf17a" was examined as "orthologues" of "fgf17a". "Fgf17a" is actually the Para log of "fgf8". "Fgf24" was actually affiliated to "fgf8" as "paralog" of zebra fish. [27]. The "phylogenetic Analyses" and "gene located analyses" pointed that auto log of "fgf24" is a Para log of "fgf18a" but not a Para log of "FGF8" it means that "fgf18a" has two paralog which are fgf18b and fgf24 [28]. Fgf20a was examined as zebra fish orthologues of "fgf20". However, further studied shows that orthologues of "fgf20b and fgf20a" are paralog of "fgf20b" [29]. At human Fgf 18 locus a gene order was ascertained. Their gene order was actually conserved. While the same "conserved" gene order was also observed at zebra fish fgf18b [30]. However, the same gene order was also studied at fgf 24 loci. They all were latently yielded from same ancestor gene [31]. They were yielded by the duplication of genes. "FGF18" and "FGF-24" were

nearly placed to each other on "chromosome 14". According to "phylogenetic" analysis, human "fgf 18" is more nearly affiliated to zebra fish "fgf 24". From the above analyses shows that fgf 24 is paralog of "fgf-18a" [32]. It is also discussed above that they are latently yielded by the process of local gene duplication [33]. While On the contrary, it is ambiguous from the gene location analyses that which of them "fgf18a or Fgf18b" is a zebra-fish "orthologues" of the "human-fgf18" [34-36]. fgf10a, fgf16, and fgf24 are needed for the fin bud to shape. Fgf19 is needed for forebrain development. Fgf20a is needed for starting fin regeneration. Fgf21 is needed for hematopoietic. As zebra fish undeveloped organisms are little, the preparation and resulting early stage advancement happen remotely, and the undeveloped improvement is fast.

Conclusion

The current study indicates that Human "fgf 18" is more closely related to zebra fish "fgf 24" according to "phylogenetic" study. According to the results of the above studies, fgf 24 is a paralog of "fgf-18a". They are also latently generated by the method of local gene duplication. It is unclear whether "fgf18a or Fgf18b" is a zebra-fish "orthologue" of the "human-fgf18". fgf10a, fgf16, and fgf24 are needed for the fin bud to form based on gene location analyses. Fgf19 is required for the development of the forebrain. Since the undeveloped species of zebra fish are small, the planning and subsequent early stage development take place remotely, and the undeveloped development is rapid.

References

- [1] Gospodarowicz D, Jones KL, Sato G (1974). Purification of a growth factor for ovarian cells from bovine pituitary glands. *Proc Natl Acad Sci USA* 71:2295-99.
- [2] Ornitz DM, Itoh N (2001). Fibroblast growth factors. *Genome Biol.* 2:1-2.
- [3] Goldfarb M (1996). Functions of fibroblast growth factors in vertebrate development. *Cytokine Growth Factor Rev* 7:311-325.
- [4] Eswarakumar VP, Lax I, Schlessinger J (2005). Cellular signaling by fibroblast growth factor receptors. *Cytokine Growth Factor Rev* 16:139-49.
- [5] Powers CJ, McLeskey SW, Wellstein A (2000). Fibroblast growth factors, their receptors and signaling. *Endocr Relat Cancer* 7:165-97.
- [6] Schlessinger J (2000). Cell signaling by receptor tyrosine kinases. *Cell* 103:211-25.
- [7] Sleeman M, Fraser J, McDonald M, et al. (2001). Identification of a new fibroblast growth factor receptor, FGFR5. *Gene* 271:171-82.
- [8] Lee PL, Johnson DE, Cousens LS, et al. (1989). Purification and complementary DNA cloning of a receptor for basic fibroblast growth factor. *Science* 245:57-60.

- [9] Itoh N, Ornitz DM. (2004). Evolution of the Fgf and Fgf gene families. *Trends Genet* 20:563–69.
- [10] Olsen SK, Garbi M, Zampieri N, et al. (2003). Fibroblast growth factor (FGF) homologous factors share structural but not functional homology with FGFs. *J Biol Chem*. 278:34226-36.
- [11] Fu L, John LM, Adams SH, et al. (2004). Fibroblast growth factor 19 increases metabolic rate and reverse dietary and leptin-deficient diabetes. *Endocrinology* 145:2594-603.
- [12] Kharitonov A, Shiyanova TL, Koester A et al. (2005). FGF-21 as a novel metabolic regulator. *J Clin Invest* 115:1627-35.
- [13] Razzaque MS, Lanske B. (2007). The emerging role of the fibroblast growth factor-23-klotho axis in renal regulation of phosphate homeostasis. *J Endocrinol*. 194:1–10.
- [14] Tomlinson E, Fu L, John L, et al. (2002). Transgenic mice expressing human fibroblast growth factor-19 display increased metabolic rate and decreased adiposity. *Endocrinology*. 143:1741-47.
- [15] Lin X, Buff EM, Perrimon N, et al. (1999). Heparin sulfate proteoglycan is essential for FGF receptor signaling during *Drosophila* embryonic development. *Development* 126:3715–23.
- [16] Ornitz DM, Yayon A, Flanagan JG, et al. (1992). Heparin is required for cell-free binding of basic fibroblast growth factor to a soluble receptor and for mutagenesis in whole cells. *Mol Cell Biol* 12:240–47.
- [17] Kolpakova E, Stenmark H, Klingenberg O, et al. (1998). Cloning of an intracellular protein that binds selectively to mitogenic acidic fibroblast growth factor. *Biochemical Journal* 336:213-22.
- [18] Popovici C, Roubin R, Coulier F, et al. (2005). An evolutionary history of the FGF super family. *Bioassays*. 27:849-57.
- [19] N. Itoh (2007). The Fgf families in humans, mice, and zebra fish: their evolutionary processes and roles in development, metabolism, and disease. *Biol. Pharm. Bull.* 30:1819-25.
- [20] Thisse B, Thisse C (2005). Functions and regulations of fibroblast growth factor signaling during embryonic development. *Dev Biol* 287:390–402.
- [21] Goldfarb M (2005). Fibroblast growth factor homologous factors: evolution, structure, and function. *Cytokine Growth Factor Rev* 16:215–20.
- [22] Shin JT, Fishman MC (2002). From zebra fish to human: modular medical models. *Annu Rev Genomics Hum Genet* 3:311–40.
- [23] Beis D, Stainier DY (2006). In vivo cell biology: following the zebra fish trend. *Trends Cell Biol* 16:105–112.
- [24] Patton EE, Zon LI (2001). The art and design of genetic screens: zebra fish. *Nat Rev Genet* 2:956–66.
- [25] Stern HM, Zon LI. (2003). Cancer genetics and drug discovery in the zebra fish. *Nat Rev Cancer* 3:533–539.
- [26] Zon LI, Peterson RT (2005). In vivo drug discovery in the zebra fish. *Nat Rev Drug Discov* 4:35–44.
- [27] Kiefer P, Strahle U, Dickson C (1996). The zebra fish Fgf-3 gene: cDNA sequence, transcript structure and genomic organization. *Gene* 168:211–15.
- [28] Furthauer M, Thisse C, Thisse B (1997). A role for FGF-8 in the dorsoventral patterning of the zebra fish gastrula. *Development* 124:4253–64.
- [29] Horton AC, Mahadevan NR, Ruvinsky I, (2003). Phylogenetic analyses alone are insufficient to determine whether genome duplication(s) occurred during early vertebrate evolution. *J Exp Zool B Mol Dev Evol* 299:41–53.
- [30] Nusslein-Volhard C, Dahm R, editors. *Zebra fish*. Oxford University Press; 2002
- [31] Scholpp S, Groth C, Lohs C (2004). Zebra fish fgfr1 is a member of the fgf8 syn expression group and is required for fgf8 signaling at the mid- brain-hindbrain boundary. *Dev Genes Evol* 214: 285–95.
- [32] Tonou-Fujimori N, Takahashi M, Onodera H et al. (2002). Expression of the FGF receptor 2 gene (fgfr2) during embryogenesis in the zebra fish *Danio rerio*. *Mech Dev* 119:S173–S78.
- [33] Sleptsova-Friedrich I, Li Y, Emelyanov A, et al. (2001). Fgfr3 and regionalization of anterior neural tube in zebra fish. *Mech Dev* 102:213–17.
- [34] Thisse B, Thisse C, Weston JA (1995). Novel FGF receptor (Z- FGFR4) is dynamically expressed in mesoderm and neurectoderm during early zebra fish embryogenesis. *Dev Dyn* 203:377–91.
- [35] Nasevicius A, Ekker SC (2000). Effective targeted gene 'knockdown' in zebra fish. *Nat Genet* 26:216–20.
- [36] Picker A, Brennan C, Reifers F, et al. (1999). Requirement for the zebra fish mid-hind- brain boundary in midbrain polarization, mapping and confinement of the retinotectal projection. *Development* 126:2967–78.